

Marking and Identifying Free-Roaming Dogs and Cats

Valerie Benka & Susan Getty





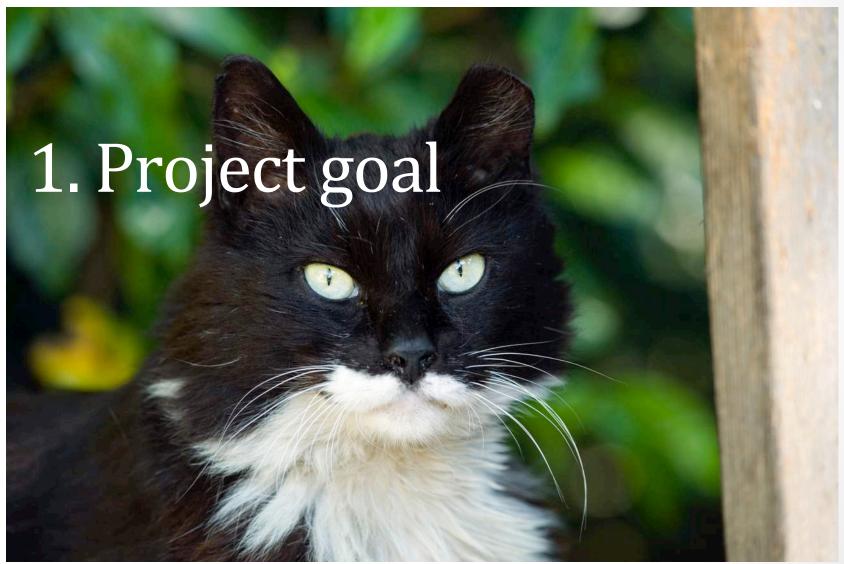


ACC&D 6th International Symposium • July 24, 2018

Presentation Outline

- Project goal
 - 1. Think Tank
 - 2. Marking criteria
- 2. Cornell partnership to develop ear tags
- 3. Pilot/field studies
 - 1. Sheltered dogs in Romania (Summer 2015)
 - Study in pet cats in Illinois (March 2016)
 - 3. Study in free-roaming owned dogs in Kenya (September 2016)
 - 4. Next steps; 2nd study in cats in Illinois
- 4. Microneedle patch







Project Goal: To identify a new method, or improve upon an existing method, to mark animals as non-surgically sterilized or contracepted.

Think tank

- Broad expertise
- Review current marking across species and contexts
- Establish criteria for nonsurgical marking of dogs and cats



Identifying & Prioritizing Marking Methods for Non-Surgically Sterilized Cats & Dogs Scientific Think Tank

> May 16-18, 2013 Phoenix, AZ

Overview

On May 16-18, 2013, the Alliance for Contraception in Cats & Dogs (ACC&D) convened a Think Tank on identifying and prioritizing marking methods for non-surgically sterilized cats and dogs. The meeting was held in Phoenix, Arizon, with financial support flow of Personart Charities, ACC&D undestrook this initiative in response to the growing need for a means to identify animals who have been treated with a non-surgical sterilant or long-term contraceptive. The Think Tank focused on cats and dogs, particularly free-roaming populations, who have undergone non-surgical interventions. However, the use is not limited to these animals—improved marking methods could have benefits for surgically sterilized animals as well.

The mission of ACC&D is to expedite the successful introduction and to support the distribution and promotion of non-surgical methods to sterilize cats and dogs. ACC&D vision is to reduce animal death and suffering worldwide by enhancing the tools available to humanely control their populations. Non-surgical approaches can be less expensive, less labor-intensive, and require shorter recovery times than surgery, allowing more animals to be treated quickly and safely.

We gratefully acknowledge PetSmart Charities for sponsoring this Think Tank.



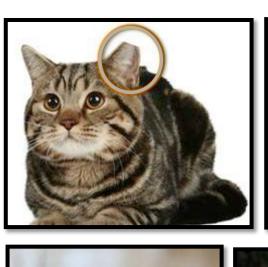
This was ACC&D's fifth scientific Think Tank. Prior Think Tanks have addressed:

- Population modeling as a tool to guide the design and implementation of contraceptive approaches to best achieve stabilization or reduction of population size
- Delivery technologies that could be used to precisely control administration of slow-release, timed-release, or multi-dose treatments
- Methods to improve immunocontraceptive vaccines for sterilization of dogs and cats
- Gene silencing and immunocontraception as research areas with promise for achieving the goal of a non-surgical sterilant

The marking methods Think Tank sought to identify the most promising near-term and long-term methods to mark animals the near-term and long-term methods to mark animals the near-term and long-term methods to mark animals the near-term and the nea

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Current Animal Marking Methods







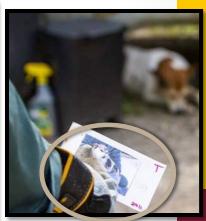












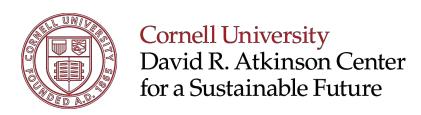
Marking Criteria: Minimum and Ideal

Criteria		Minimum	Ideal	
Visibility		~ 12 ft	>25 ft	
Permanence		>3 years	Life of animal	
Behavioral Impact (i.e., interference with normal behavior, other animals, or humans)		None	None	
Application	Time required	<10 minutes	5 seconds	
	Training required	Little	None	
	Humane/pain level	No anesthesia, pain controllable/very brief	No anesthesia, no pain	
	Cost per application	<\$10	<\$1	
Info Retrieval	Ease of Retrieval	Visual reading or simple device	Visual + data capture	
	Quantity of information	Treated (yes/no)	Type/date(s) of treatment, other info	
	Info retrieval device cost	<\$50	\$0	





R&D partners and funders











Cornell University
College of Human Ecology
Fiber Science & Apparel Design

Key Cornell players:

- PI: Margaret Frey, PhD (Fiber Science)
- Co-Inv: Elizabeth Berliner, DVM (CVM)
- Co-Inv: Edwin Kan, PhD (Engineering)
- DVM student: Eloïse Cucui

Partners and volunteers:

- Kaos Softwear, Portland, OR
- Dr. Karl Citek, Pacific University College of Optometry
- Associaţia de Protecţie a Animalelor "Milioane de Prienteni", Romania
- Dr. Kevin McGowan, Cornell Ornithology Lab
- David Buffington, Glen Raven Custom Fabrics
- Gene Pancheri, Proctor & Gamble (ret).

Development – multiple expertise

Tag Material

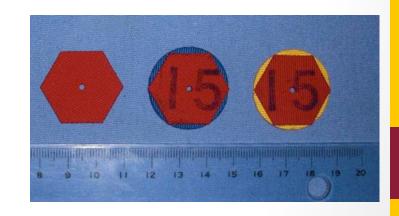


Application

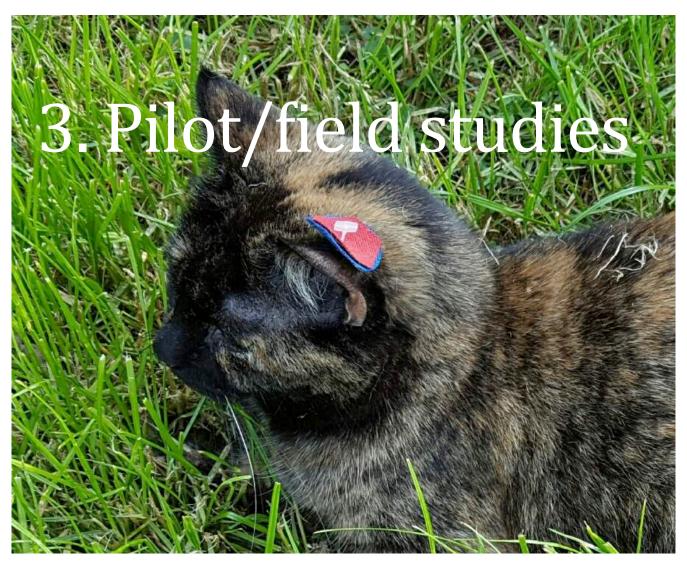


Coding/Visibility













Animals & Methods

- 54 dogs of both sexes and varied ages and types
- Tag applied under anesthesia for s/n surgery
- Observations on days 0-8, 10, 12, 14, 21, 28, etc.





Photo credits: Eloïse Cucui



- Applicator works well in anesthetized animals
- Tags do not cause infection or pain in animals treated with antibiotics and analgesics
- Tag visibility is not always ideal;
 RFID beneficial



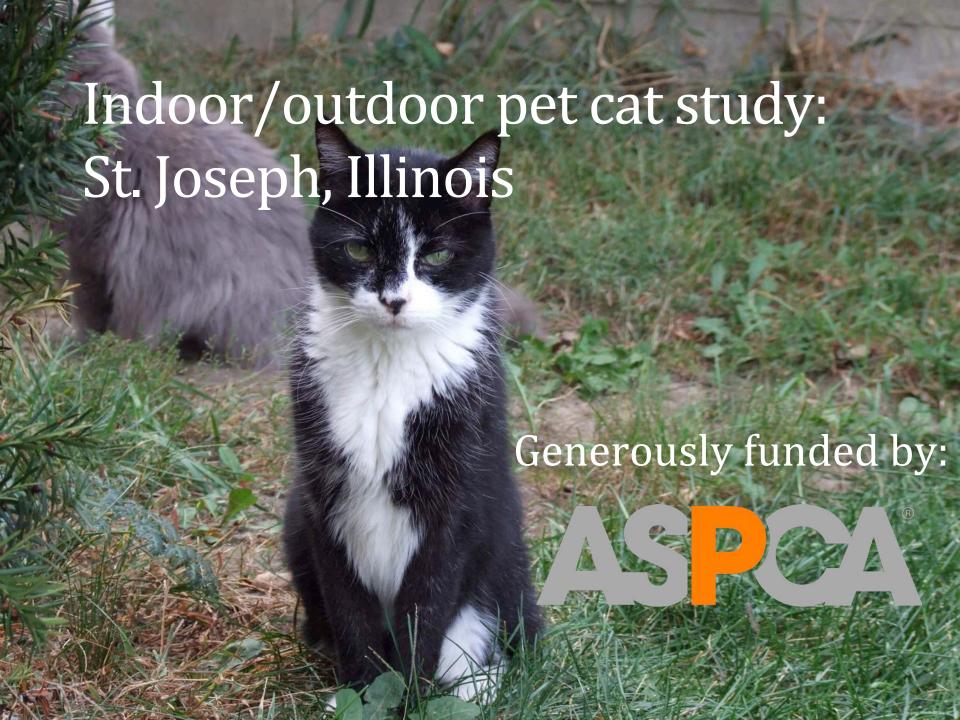


Results

- Tag loss was low but significant; 5 of 54 tags (9.25%) failed over 643 dog days = 0.008 tags/dog day
- Tags might not work for closely confined and/or teething puppies
- Multiple outstanding questions for future field studies – not least tolerance of application and durability









Animals & Methods

- 9 indoor/outdoor pet cats belonging to Amy Fischer
- Tag applied under anesthesia for routine or Grade 1 periodontal treatments
- Same monitoring protocol as in Romania



Results: Ears

- Cats developed slight scabbing at point of application—not serious or bothersome
- Skin underneath tag was healthy







Results: Behavior

- All but one cat exhibited normal behavior throughout.
- One female initially exhibited ear flicking and head shaking, which stopped within a few days.
- No change in non-tagged cats' behavior toward tagged cats.





Results: Tag Loss

- 3/9 cats lost tags (D19, 30, 119)
 - All tags failed at the fastener
 - No damage to ear
- 2/9 cats have tags after 28 months
- 4/9 removed for reasons unrelated to tag failure

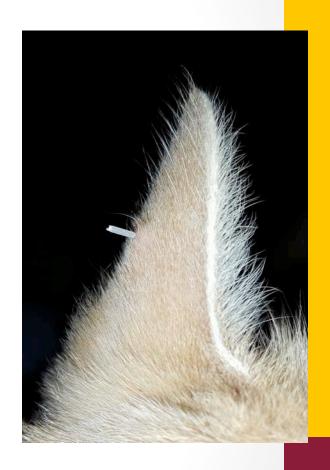


Photo credit: Amy Fischer





Animals & Methods

- Objectives: evaluate 1) practicality and humaneness of application in conscious dogs, 2) tag performance
- Study population: >100 free-roaming owned dogs with owner permission to tag.
- Selected calm, stable dogs who did not respond to handling or other medical treatments.
- Used ethyl chloride topical anesthetic spray





Results

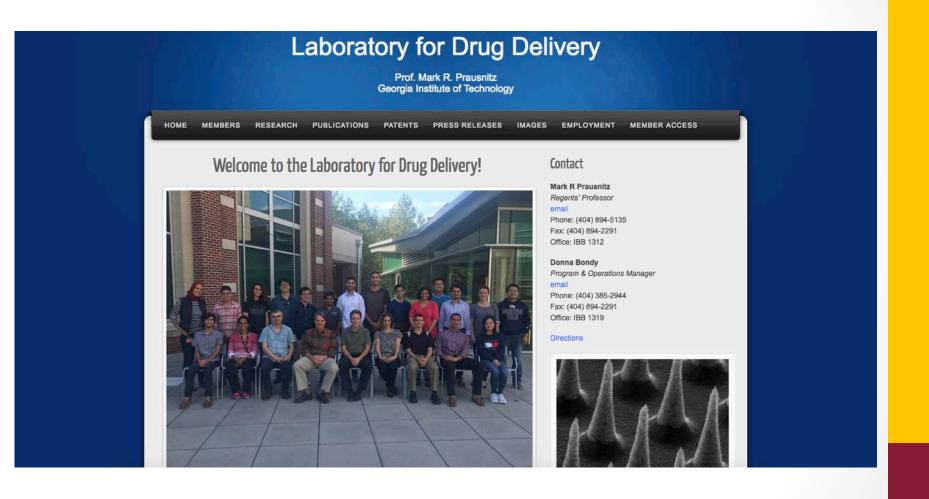
- Dogs struggled and vocalized during ethyl chloride and tag application
 - Indicators of pain
 - Ethyl chloride did not have adequate anesthetic effect
- Attempted tagging in 6 dogs
 - Four successful, two aborted
- Study halted on animal welfare grounds
- Pain/distress was very transient; dogs behaved normally within minutes post-procedure
- Tag loss: 3 dogs followed, all had lost tags by day 16



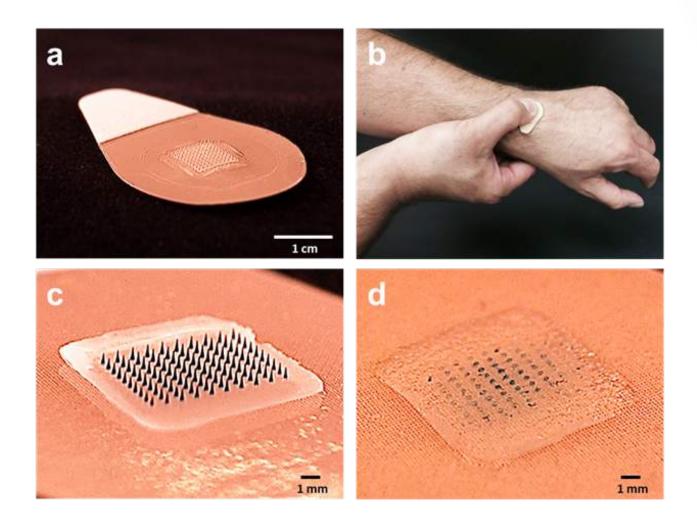
Marking Conclusions

- Suspend further lab or controlled field studies of prototype ear tag in dogs due to combined:
 - Indications of pain
 - Limited anesthetic options for field use
 - Tag loss
- Still a possibility for cats?

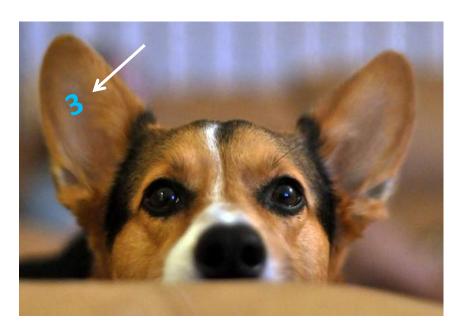
4. Partnership with Dr. Mark Prausnitz of Georgia Tech

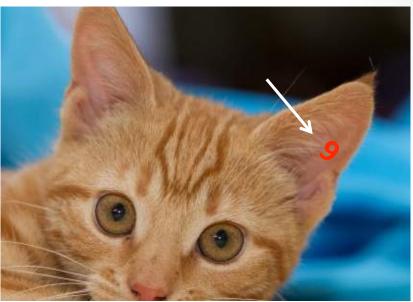


What is a microneedle patch and how does it work?

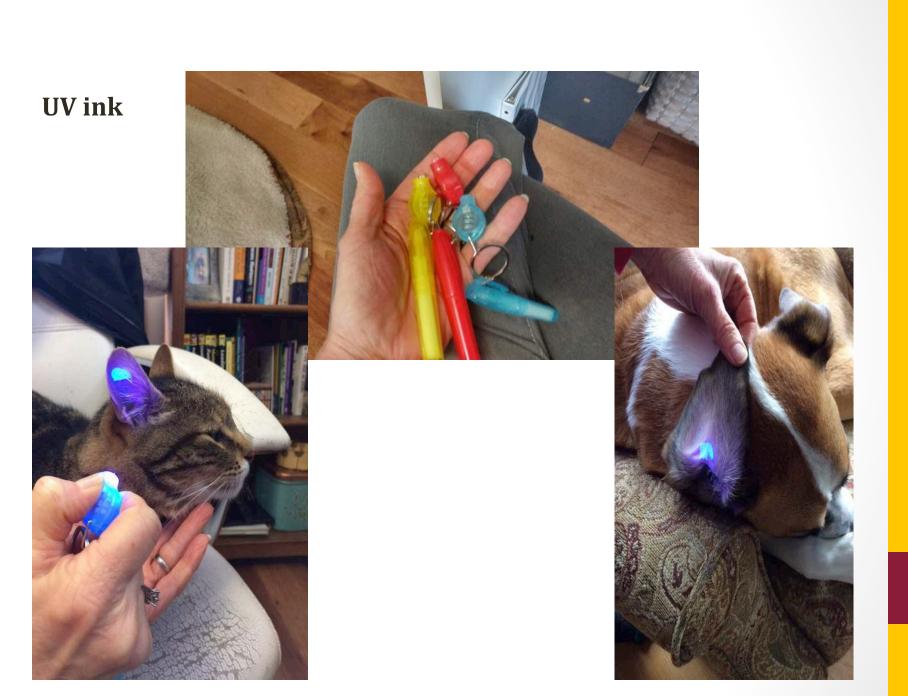


How would this work on dogs and cats?





- Color indicates gender
- Number indicates year



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Contents lists available at ScienceDirect

Journal of Controlled Release





Rabies vaccination in dogs using a dissolving microneedle patch

Jaya M. Arya a, Kristopher Dewitt b, Maya Scott-Garrard b, Yu-Wei Chiang b, Mark R. Prausnitz a,*



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ABSTRACT

Because humans get rabies primarily through dog bites, stray dog population control and mass or mandatory vaccination of domestic dogs and other animals has virtually eliminated human rabies in industrialized countries. However, thousands of people in developing countries die of rabies each year due to the inability to control dog populations and implement mass vaccination because of financial, logistical and other challenges. The availability of an easier-to-administer and more cost-effective vaccine may help to address some of these issues. Here, we propose the use of dissolving microneedle patches for simple and potentially cost-effective rabies vaccination, and assess the safety and immunogenicity of microneedle patch vaccination using a rabies DNA vaccine in dogs. The vaccine was stable upon formulation and storage for at least 3 weeks at 4 °C in a microneedle patch. For vaccination, the patches were applied to the inner ear by hand without an applicator. Microneedle patches were well tolerated in the skin, with mild erythema, minimal wheal formation and complete resolution of skin reactions within 7 days, and generated no systemic adverse events. Microneedle patches were at least as immunogenic as intramuscular injection at the same dose, as demonstrated by similar serum neutralizing antibody titers. A ten-fold lower vaccine dose administered by microneedle patch generated a weaker immune response compared to full-dose intramuscular vaccination. We conclude that dissolving microneedle patches may provide an innovative approach to mass vaccination of dogs.

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Table 1
Tolerance of dogs to vaccination by IM injection and microneedle patch¹.

	Prime (day 0)	Boost (day 28)]
Intramuscular injection 50 µg	60%	O %	Dogs were
Microneedle patch 50 μg	O %	O%	tolerant
Microneedle patch 5 μg	O %	20%	Dogs were intolerant
Placebo microneedle patch	O%	O%	

Dogs were considered intolerant of injection if they vocalized, withdrew or tried to bite upon injection.

Table 2 Number and type of local injection site reactions¹.

	Erythema	Wheal formation	Swelling	Pain upon palpation	Ulceration	Skin
IM 50 μg	O 0%	0.0%	O 0%	OOK	O 0%	reactions absent Skin reactions
MN 50 μg	● 100%	₾20%	O 0%	O 0%	0%	
MN 5 μg	●100%	40%	O 0%	O 0%	0%	
MN Placebo	3 50%	1 50%	O 0%	00%	O 0%	

¹ The dogs were observed for local injection site reactions on the day of the vaccination, daily for the first three days following each vaccination and intermittently for any dogs with reactions persisting for more than three days. This table reports the cumulative percentage of dogs with injection site reactions after both vaccinations.

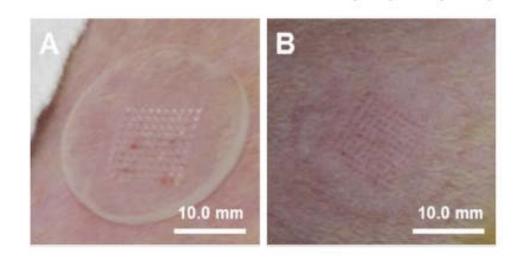
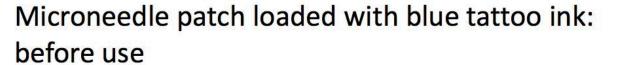


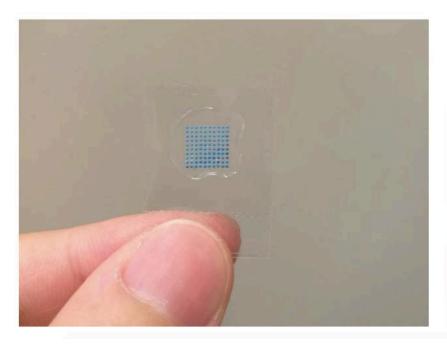
Fig. 5. Dog ear during and after microneedle patch vaccination in vivo. Microneedle patches were applied onto dog ears with hair removed, left on the skin for 15 min and then removed. (A) Microneedle patch applied to skin. (B) Same section of skin immediately imaged after microneedle patch application and removal showing a faint grid where microneedles inserted and slight skin erythema.

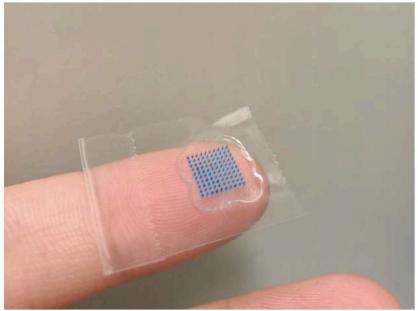
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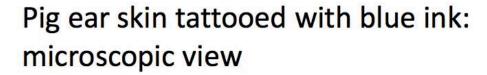
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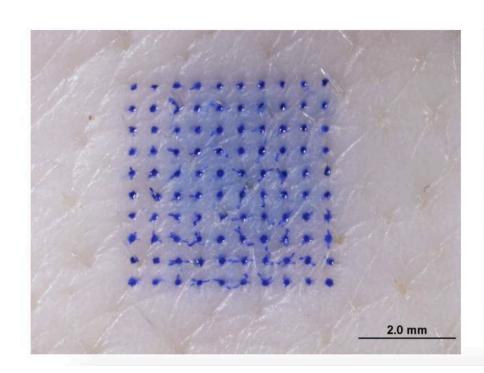


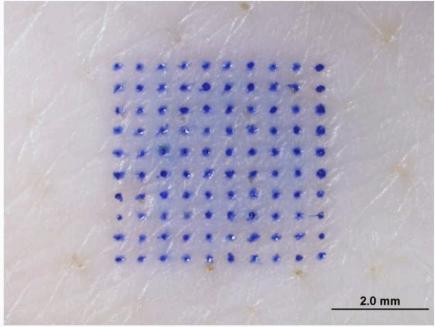




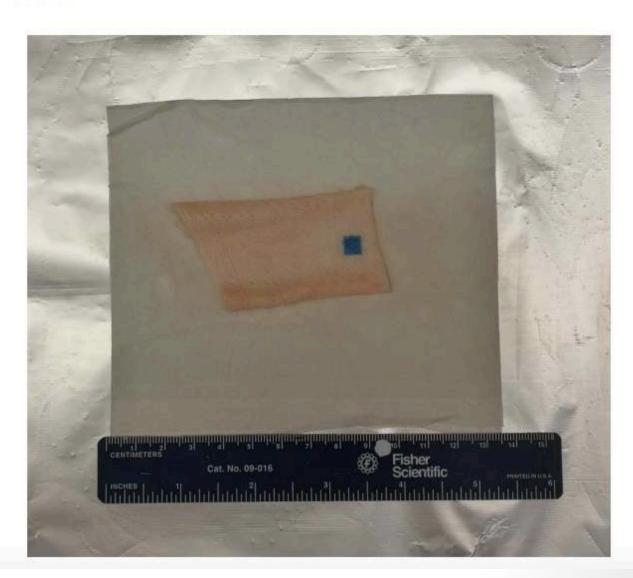








Pig ear skin tattooed with blue ink: macroscopic view



Pig ear skin tattooed with blue ink: viewed at a distance

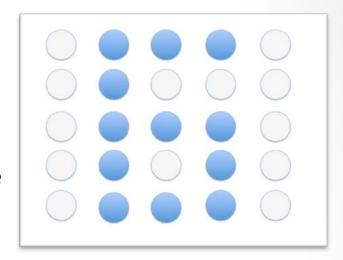


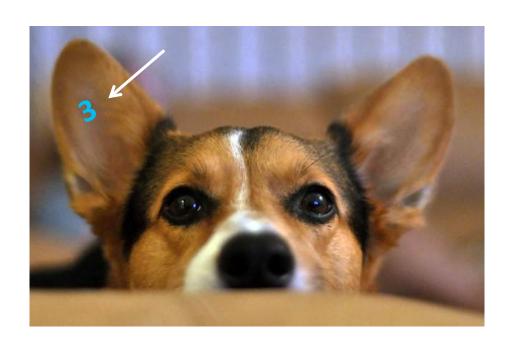
Pig ear skin tattooed with blue ink: viewed at a distance



Next Steps:

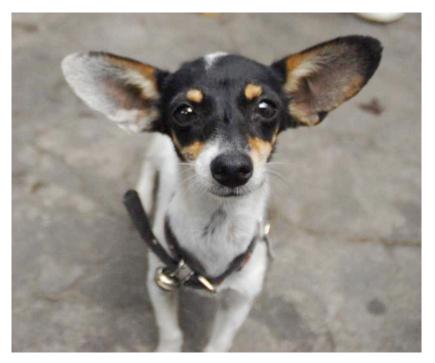
- ink patterned in numbers
- UV ink
- Patch with shorter application time
- Patch with biodegradable adhesive





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Potential problems

We welcome your feedback!

- For those of you working in the field, what are your thoughts?
 - When would this work, and when wouldn't it work?
 - If we couldn't get application to a matter of seconds, would it work to press the patch on, have it be held on with a sticky, biodegradable material that falls off eventually?

